NEWLY IDENTIFIED MESSAGE FOR RESCUE AND REPAIR FROM NECROTIC CELL: BIOLOGY AND CLINICAL RELEVANCE OF "ENDOKINE, HMGB1"

Ikuro Maruyama, Takashi Ito and Teruto Hashiguchi

Abstract Responses to stimuli in cellar level are diverse and such hierarchical as secretion of stored factors, synthesis of lipid mediators and protein synthesis through genomic transcription. However, how can the cells respond in the case of necrosis? Recently a characteristic intranuclear protein, high-mobility group box 1 protein (HMGB1) is released from necrotic cells. The protein is an abundant nuclear protein with a dual function both inside and outside the cells. In physiological state, HMGB1 is present in the nucleus, and binds to DNA, playing a variety of crucial functions, including transcription and keeping the characteristic DNA architecture. However, the protein is released to extracellular space from most of necrotic cells, activated macrophages and dendritic cells. Out of the cells, HMGB1 acts as a signal of tissue damage and can promote inflammation, immune responses, and results tissue regeneration. During sepsis and/or disseminated intravascular coagulation (DIC), however, massive accumulation of HMGB1 in the systemic circulation will cause multiple organ failure (MOF) and subsequent lethal outcome. Thus HMGB1 in the systemic circulation has been considered as a lethal mediator of sepsis, and a promising therapeutic target for sepsis. Recently we identified that thrombomodulin (TM), a natural anticoagulant glycoprotein expressed on the surface of endothelial cells, plays an important role in sequestering HMGB1. TM may prevent HMGB1 from reaching remote organs, thereby restricting the spectrum of HMGB1 action in the site of injury. Here we review recent progress made in defining the physiological and pathological roles of HMGB1 and therapeutic strategies aimed at blocking circulatory HMGB1.

Hirosaki Med. J. 59, Supplement : S1-S11, 2007

Key words: sepsis; disseminated intravascular coagulation (DIC); thrombomodulin; alarmin; highmobility group box 1 (HMGB1)

Introduction

During evolution, multi-organ lives with closed circulatory system have developed defense mechanisms to counteract life-threatening events, such as infection, trauma injury, and hemorrhage. To initiate and accomplish appropriate and rapid protective responses, our defense systems need to recognize potentially life-threatening events. So far, two theories have been proposed to explain how our defense systems detect such threatening: stranger hypothesis and danger hypothesis^{1,2)}. Immune cells possess molecular microdetectors, called pattern recognition receptors (PRRs), that recognize common molecular patterns of invading pathogens^{3,4)}. Such pathogen-associated molecular patterns (PAMPs) include component of bacterial cell walls, flagellar proteins, and viral nucleic acids, and immunocompetent cells can detect PAMPs as life-threatening strangers using PRRs⁴⁻⁶⁾. Another strategy to detect life-threatening events is targeted on tissue damage. During trauma or microbial attack, the resulting cellular stress or tissue damage alerts our defense systems. In this case, instead of recognizing PAMPs, immunocompetent cells recognize molecules that are normally found only inside cells, unless released by damage. These

Phone: +81-99-275-5437, Fax: +81-99-275-2629, E-mail: rinken@m3.kufm.kagoshima-u.ac.jp

Department of Laboratory and Vascular Medicine, Kagoshima University Graduate School of Medical and Dental Sciences, 8-35-1, Sakuragaoka, Kagoshima, Japan

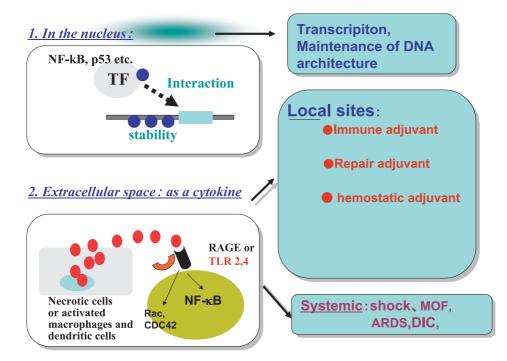


Figure 1 Intranuclear HMGB1 is release into outer cellular space by two manners. Nuclear protein HMGB1 is passively released from most of necrotic cells. The protein is also actively released from activated macrophages and dendritic cells. Extracellular HMGB1 acts as an adjuvant in immune, wound healing and hemostatic reaction in localized damaged tissue. However, circulatory systemic HMGB1 may metastasize inflammation and show remote effect resulting DIC, MOF and shock.

intracellular molecules include uric acid, RNA, high-mobility group box 1 protein (HMGB1), heatshock proteins (HSPs), and S100 proteins, and the term damage-associated molecular patterns (DAMPs) or "alarmin" is proposed to categorize such endogenous molecules that signal tissue damage^{2,7-9}. Thus, our defense systems scan for sings of both stranger and danger, and initiate protective responses, such as inflammation, immune response, hemostasis, and tissue repair.

Extracellular HMGB1 is a message for rescue from the necrotic cells

HMGB1 was previously thought to function only as a nuclear factor that enhances transcription. This protein is expressed in almost all eukaryotic cells, and is highly conserved through evolution^{7,10}. HMGB1 is highly mobile and can shuttle between the nucleus and the cytoplasm¹¹. HMGB1 binds to the minor groove of DNA, without sequence specificity, which causes a conformational change in the DNA that promotes its interaction with transcription factors such as NF- κ B, steroid hormone receptors, and glucocorticoid receptors^{12,13)}. HMGB1 gene knock out mice die shortly after birth because of, at least in part, hypoglycemia caused by deficient glucocorticoid receptor function¹⁴⁾. Besides its intracellular role, HMGB1 can be released into the extracellular milieu and plays quite distinct roles. There are two ways in which HMGB1 can be released: a passive release from necrotic cells and an active secretion from immunocompetent cells (Figure 1) ¹⁵⁻¹⁷⁾.

Normally HMGB1 is bound loosely to chromatin, so if cells become necrotic or leaky, then HMGB1 diffuses out into the extracellular milieu¹⁷⁾. Importantly, apoptotic cells do not release HMGB1 even after undergoing secondary necrosis and partial autolysis¹⁷⁾. Thus, cells

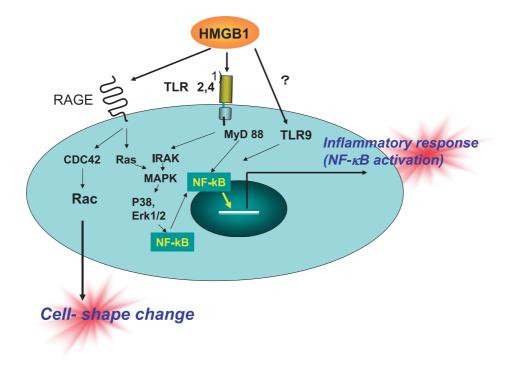


Figure 2 HMGB1 and its multiple receptor system. Extracellular HMGB1 acts on RAGE and Toll-like receptor-2 and -4, and results activations of NF κ B and CDC2, Rac. These actions of HMGB1 play a crucial role in the development in DIC/MOF and septic shock.

undergoing apoptosis are programmed to withhold the signal, thereby preventing the inflammatory process during apoptosis. Several cell types, such as inflammatory cells, possess the ability to secrete HMGB1 actively, and thus produce a danger signal without dying. The active secretion depends on a relocalization of HMGB1 from the nucleus to specific cytoplasmic organelles, the secretory lysosomes¹⁶. Inflammatory stimuli lead to hyperacetylation of HMGB1, which causes an accumulation of HMGB1 in the cytoplasm and blocks a reentry to the nuclear compartment^{18,19}. Cytoplasmic HMGB1 is taken up by secretory lysosomes, and then the HMGB1-containing secretory lysosomes are fused with the plasma membrane and are secreted, at least in part, in a specific ATP-binding cassette (ABC) transporterdependent manner^{9,16)}. Once released into the extracellular space, HMGB1 acts as a signal of tissue damage and can promote inflammation through specific receptors including RAGE (Receptor for advanced glycation endproducts), Toll-Like receptor-2 (TLR2) and TLR4, and possibly other receptors²⁰⁻²⁵⁾ (Figure 2).

HMGB1 interacts with RAGE expressed on endothelial cells, inducing endothelial cell activation, with increased expression of vascular cell adhesion molecule 1 (VCAM1), intercellular adhesion molecule 1 (ICAM1), and E-selectin^{26,27)}. In addition, HMGB1 activates adhesive and migratory functions of neutrophils and monocytes in a AGER-dependent manner^{28,29)}. Engagement of RAGE therefore increases adhesion of neutrophils and monocytes to endothelial cells, and promotes the recruitment of leukocytes across endothelial barriers. HMGB1 also acts on leukocytes, inducing the expression of proinflammatory cytokines, such as tumor necrosis factor-a (TNF-a), interleukin-1 (IL-1), IL-6, and type I interferon (IFN)^{15,30,31)}. In this regard, HMGB1 and PAMPs synergistically activate leukocytes, and it may contribute both to our defense systems against pathogen and to autoimmune pathogenesis²⁵⁾. Necrotic cells lacking HMGB1 have a greatly reduced ability to elicit the production of TNF-a by neighborhood monocytes, suggesting an important role of HMGB1 in promoting inflammation during necrotic cell death¹⁷⁾. Extracellular HMGB1 also acts as an endogenous immune adjuvant. For the clonal expansion of naïve T cells, both specific antigen and co-stimulatory signals provided by the same antigen-presenting cell (APC) are required³²⁾. In bacterial infections, microbial components act as immune adjuvants and induce the expression of the co-stimulatory molecules on APCs³³⁾. However, there are some circumstances where immune responses are generated in the apparent absent of any microbial or other exogenous adjuvants. Such situations include immune responses to tumors, transplants, and possibly certain viruses. It has been postulated that in such conditions APCs can be activated by endogenous signals derived from cells that are stressed, virally infected, or killed necrotically, but not by healthy cells^{34,35)}. Now several actual and putative endogenous adjuvants, such as HSPs, uric acid, and HMGB1, have been identified, and there are others whose identities are not yet known (Figure 1b)³⁶⁻³⁸⁾. In addition to its roles in inflammation and immune responses, extracellular HMGB1 has the ability to promote tissue repair. Compared to healthy muscle, dystrophic muscle contains more HMGB1, and exogenously added HMGB1 induces migration and proliferation of vessel-associated stem cells³⁹⁾. After myocardial infarction, exogenously added HMGB1 induces proliferation and differentiation of cardiac progenitor cells, thereby promoting cardiac regeneration⁴⁰. Furthermore, cardiac performance in HMGB1-treated mice is significant improved compared to control mice, suggesting that HMGB1 can induce functionally relevant myocardial regeneration following myocardial infarction⁴⁰. Although the role of endogenous HMGB1 in tissue regeneration remains to be clarified, exogenously added HMGB1 may be used to promote tissue regeneration. Taken together, HMGB1 released from damaged or activated cells seems to orchestrate post-injury responses: inflammation, immune responses, and subsequent tissue regeneration.

HMGB1 as a lethal mediator in sepsis and/or DIC

Sepsis is a life-threatening disorder that results from systemic inflammatory and coagulatory responses to infection⁴¹⁾. Hyperactivation of the inflammatory system is the most important feature of sepsis, and has been the most common target of therapeutic strategies. So far, diverse therapies directed against proinflammatory mediators such as TNF- α and IL-1, have revealed dramatic effects in animal models of sepsis^{42,43)}. However, in humans, most of these strategies have not improved survival of septic patients $^{44,45)}$. It is in part because the classical proinflammatory mediators such as TNF- α and IL-1 are released within minutes after PAMPs exposure, then, even a minimal delay in treatment may result in treatment failure. HMGB1 is one promising therapeutic target for sepsis. In sepsis and/or disseminated intravascular coagulation (DIC), serum HMGB1 levels are elevated in both humans and animals⁴⁶⁻⁴⁸⁾ (Figure 3).

The accumulation of HMGB1 in the systemic circulation occurs considerably later than that of classically early proinflammatory mediators such as TNF-a and IL-1^{46,49}, and this delayed kinetics of HMGB1 makes it an attractive therapeutic target with a wider window of opportunity for the treatment of sepsis⁵⁰. Indeed, blockade of HMGB1, even at later time points after onset of infection, has been shown to rescue animals from lethal sepsis^{46,51-53}. Although data on higher species are not yet available, HMGB1 antagonism provide hope for new approaches that will be

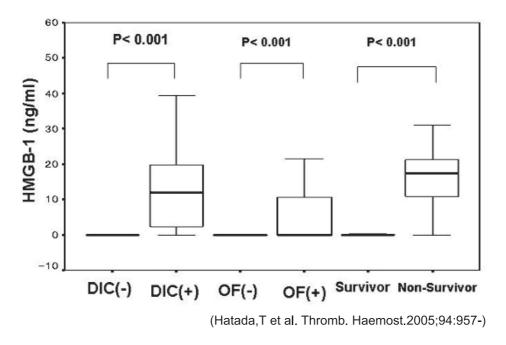


Figure 3 Elevated HMGB1 level in the plasma from the DIC patients. Among various infectious diseases, plasma HMGB1 was increased in the plasma from the patients complicated with DIC, MOF and lethality.

therapeutically effective in humans with sepsis. The mechanisms by which HMGB1 exerts its lethal effect in sepsis are not fully determined and seem to be multifactorial. Proinflammatory activity of HMGB1 is one possible mechanism. Extracellular HMGB1 triggers inflammation, and HMGB1 plays an important role in the pathogenesis of inflammatory diseases, including acute inflammatory lung injury, rheumatoid arthritis, and vasculitis^{30,54-58)}. Therefore, massive accumulation of HMGB1 in the systemic circulation will lead to systemic inflammatory response syndrome (SIRS), a major feature of sepsis. However, it is also reported that the roles of HMGB1 and TNF are distinguishable^{7,50)}, and the lethal effects of HMGB1 in sepsis cannot be explained by its inflammatory activity only. HMGB1 may alter the procoagulant-anticoagulant balance, with an increase in procoagulant factors and a decrease in anticoagulant factors⁵⁹⁾, and such alteration of the procoagulant-anticoagulant balance is another important aspect of sepsis⁶⁰. HMGB1 induces tissue factor expression on monocytes, and inhibited anticoagulant protein C pathway mediated by thrombin-thrombomodulin (TM) complexes⁵⁹⁾. In thrombin-induced DIC model rats, HMGB1 accelerates microvascular thrombosis with irreversible renal and respiratory failure, thereby acting as a lethal mediator⁵⁹. In patients with DIC, plasma HMGB1 levels are significantly increased and correlate with the DIC score and sepsis-related organ failure assessment (SOFA) score⁴⁸⁾. Given that DIC and concomitant multiple organ failure are strong predictors of mortality, procoagulant activity of HMGB1 may in part be responsible for its lethal effects. HMGB1 may act as a lethal mediator, by causing epithelial-cell barrier dysfunction^{61,62)}. Exposure of epithelial cells to HMGB1 causes downregulation of expression of cell surface molecules that are responsible for the tight adhesion between adjacent epithelial cells, leading to epithelial hyperpermeability. When injected animals, HMGB1 B box increases ileal mucosal permeability and promotes bacterial translocation and hepatocellular injury. HMGB1 specific antibodies confer significant protection against the development of these epithelial abnormalities and subsequent organ dysfunction in animals with endotoxemia or sepsis, indicating that this might be a mechanism for how increased HMGB1 levels cause organ dysfunction^{7,47}.

Thrombomodulin as a security guard in blood vessels

In a recent clinical trial, recombinant human soluble thrombomodulin (rhsTM) significantly improved DIC⁶³⁾. Endothelial membrane-bound TM forms a high-affinity complex with thrombin, thereby inhibiting thrombin interaction with fibrinogen and protease-activated receptor-1 (PAR-1). In addition, TM enhances thrombinmediated protein C (PC) activation by more than two orders of magnitude^{64,65)}. Like membrane-bound TM, the rhsTM, which is composed of the extracellular domain of TM, also binds to thrombin, thereby promoting PC activation. Activated PC (APC) has antiinflammatory and anti-apoptotic activities that involve binding of APC to endothelial protein C receptor (EPCR) and cleavage of PAR-1⁶⁶. APC exerts an antithrombotic effect by inactivating factors Va and VIIIa, limiting the generation of thrombin. As a result of decreased thrombin levels, the proinflammatory, procoagulant, and antifibrinolytic responses induced by thrombin are reduced^{64,65,67)}. In addition, TM itself has antiinflammatory properties in its N-terminal lectin-

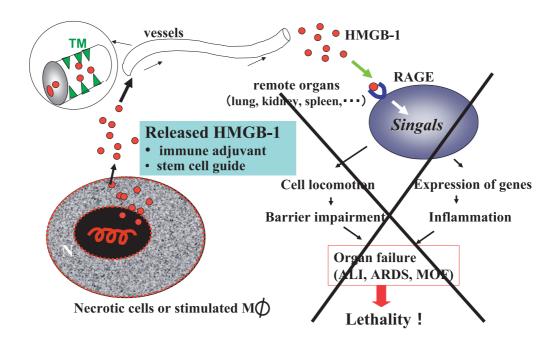


Figure 4 Endothelial thrombomodulin (TM) acts as a vascular safety guard. HMGB1 leased from necrotic cells or activated macrophages and dendritic cells acts as a adjuvant in innate immune, tissue regeneration and hemostatic system in the localized damaged tissues. Entry of the protein into the vessels and systemic circulation may be inhibited by endothelial TM by binding and subsequent degradation of the protein.

like domain⁶⁸⁾ (Figure 4).

The target molecule of TM is not restricted to thrombin. TM can bind to HMGB1 as well as thrombin, thereby dampening the inflammatory and coagulatory responses^{53,69)}. TM also dampens the generation of reactive oxygen species (ROS) (unpublished observations). Under the conditions in which injured vessel locally loses the integrity of TM-bearing cells, HMGB1 and thrombin present in the circulation will propagate inflammatory and coagulatory responses^{27,70}. However, once an adjacent portion of the vessel wall with intact endothelial cells is encountered, TM-bearing cells can sequester HMGB1 and thrombin, therefore preventing them from reaching remote organs (Figure 2). Thus, TM acts as a security guard in blood vessels. Since the expression of endothelial TM is systemically downregulated during septic conditions⁷¹⁾, replacement with rhsTM will offer therapeutic value in sepsis and/or DIC.

Concluding remarks

In the canonical model of sepsis, PAMPs are considered to be responsible for the development of sepsis. However, recent investigations suggest the importance of endogenous molecules as amplifiers of signals in the pathogenesis of sepsis^{72,73)}. Defining their functions during sepsis in greater detail will be necessary for the design of new therapeutic strategies in clinical sepsis.

Acknowledgements

We thank all our colleagues who helped us through open discussion to conceive ideas here, S. Yamada, K. Takenouchi, Y. Oyama, K. Kikuchi, Y. Morimoto, K.K. Biswas, N. Arimura, S. Arimura, N. Miura, B. Shrestha, X. Meng, S. Tancharoen, Y. Nawa, T. Shimizu, and T. Uchimura. This work was supported by the Research Grant from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (Grants-in-Aid 18791341) (T. I.) and by the Health and Labour Sciences Research Grant from the Ministry of Health, Labour and Welfare (I. M.).

References

- 1)Heath WR, Carbone FR. Immunology: dangerous liaisons. Nature 2003;425:460-1.
- 2) Bianchi ME: DAMPs, PAMPs and alarmins: all we need to know about danger. J Leukoc Biol 2007;81: 1-5.
- 3)Janeway CA Jr. Approaching the asymptote? Evolution and revolution in immunology. Cold Spring Harb Symp Quant Biol 1989, 54 Pt 1:1-13.
- Janeway CA Jr., Medzhitov R. Innate immune recognition. Annu Rev Immunol 2002;20:197-216.
- Takeda K, Kaisho T, Akira S. Toll-like receptors. Annu Rev Immunol 2003;21:335-76.
- 6) Meylan E, Tschopp J, Karin M. Intracellular pattern recognition receptors in the host response. Nature 2006;442:39-44.
- 7)Lotze MT, Tracey KJ. High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. Nat Rev Immunol 2005;5:331-42.
- 8)Kannemeier C, Shibamiya A, Nakazawa F, Trusheim H, Ruppert C, Markart P, Song Y, Tzima E, Kennerknecht E, Niepmann M, et al. Extracellular RNA constitutes a natural procoagulant cofactor in blood coagulation. Proc Natl Acad Sci USA 2007.
- 9)Harris HE, Raucci A. Alarmin (g) news about danger: workshop on innate danger signals and HMGB1. EMBO Rep 2006;7:774-8.
- 10) Erlandsson Harris H, Andersson U. Mini-review: The nuclear protein HMGB1 as a proinflammatory mediator. Eur J Immunol 2004;34:1503-12.
- 11) Falciola L, Spada F, Calogero S, Langst G, Voit R, Grummt I, Bianchi ME. High mobility group 1 protein is not stably associated with the chromosomes of somatic cells. J Cell Biol 1997;137:19-26.
- 12) Agresti A, Lupo R, Bianchi ME, Muller S: HMGB1 interacts differentially with members of the Rel family of transcription factors. Biochem Biophys Res Commun 2003;302:421-6.

- 13) Boonyaratanakornkit V, Melvin V, Prendergast P, Altmann M, Ronfani L, Bianchi ME, Taraseviciene L, Nordeen SK, Allegretto EA, Edwards DP. High-mobility group chromatin proteins 1 and 2 functionally interact with steroid hormone receptors to enhance their DNA binding in vitro and transcriptional activity in mammalian cells. Mol Cell Biol 1998;18:4471-87.
- 14) Calogero S, Grassi F, Aguzzi A, Voigtlander T, Ferrier P, Ferrari S, Bianchi ME. The lack of chromosomal protein Hmg1 does not disrupt cell growth but causes lethal hypoglycaemia in newborn mice. Nat Genet 1999;22:276-80.
- 15) Andersson U, Wang H, Palmblad K, Aveberger AC, Bloom O, Erlandsson-Harris H, Janson A, Kokkola R, Zhang M, Yang H, et al. High mobility group 1 protein (HMG-1) stimulates proinflammatory cytokine synthesis in human monocytes. J Exp Med 2000;192:565-70.
- 16) Gardella S, Andrei C, Ferrera D, Lotti LV, Torrisi MR, Bianchi ME, Rubartelli A. The nuclear protein HMGB1 is secreted by monocytes via a non-classical, vesicle-mediated secretory pathway. EMBO Rep 2002;3:995-1001.
- 17) Scaffidi P, Misteli T, Bianchi ME. Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. Nature 2002;418:191-5.
- 18) Bonaldi T, Talamo F, Scaffidi P, Ferrera D, Porto A, Bachi A, Rubartelli A, Agresti A, Bianchi ME. Monocytic cells hyperacetylate chromatin protein HMGB1 to redirect it towards secretion. EMBO J 2003;22:5551-60.
- 19) Dumitriu IE, Baruah P, Manfredi AA, Bianchi ME, Rovere-Querini P. HMGB1: guiding immunity from within. Trends Immunol 2005;26:381-7.
- 20) Hori O, Brett J, Slattery T, Cao R, Zhang J, Chen JX, Nagashima M, Lundh ER, Vijay S, Nitecki D, et al. The receptor for advanced glycation end products (RAGE) is a cellular binding site for amphoterin. Mediation of neurite outgrowth and co-expression of rage and amphoterin in the developing nervous system. J Biol Chem 1995;270:25752-61.
- 21) Taguchi A, Blood DC, del Toro G, Canet A, Lee DC, Qu W, Tanji N, Lu Y, Lalla E, Fu C, et al. Blockade of RAGE-amphoterin signalling suppresses tumour

growth and metastases. Nature 2000;405:354-60.

- 22) Schmidt AM, Yan SD, Yan SF, Stern DM. The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. J Clin Invest 2001;108:949-55.
- 23) Park JS, Svetkauskaite D, He Q, Kim JY, Strassheim D, Ishizaka A, Abraham E. Involvement of tolllike receptors 2 and 4 in cellular activation by high mobility group box 1 protein. J Biol Chem 2004;279:7370-7.
- 24) Park JS, Gamboni-Robertson F, He Q, Svetkauskaite D, Kim JY, Strassheim D, Sohn JW, Yamada S, Maruyama I, Banerjee A, et al. High mobility group box 1 protein interacts with multiple Tolllike receptors. Am J Physiol Cell Physiol 2006;290: C917-24.
- 25) Tian J, Avalos AM, Mao SY, Chen B, Senthil K, Wu H, Parroche P, Drabic S, Golenbock D, Sirois C, et al. Toll-like receptor 9-dependent activation by DNA-containing immune complexes is mediated by HMGB1 and RAGE. Nat Immunol 2007;8:487-96.
- 26) Chavakis T, Bierhaus A, Al-Fakhri N, Schneider D, Witte S, Linn T, Nagashima M, Morser J, Arnold B, Preissner KT, et al. The pattern recognition receptor (RAGE) is a counterreceptor for leukocyte integrins: a novel pathway for inflammatory cell recruitment. J Exp Med 2003;198:1507-15.
- 27) Fiuza C, Bustin M, Talwar S, Tropea M, Gerstenberger E, Shelhamer JH, Suffredini AF. Inflammation-promoting activity of HMGB1 on human microvascular endothelial cells. Blood 2003;101:2652-60.
- 28) Orlova VV, Choi EY, Xie C, Chavakis E, Bierhaus A, Ihanus E, Ballantyne CM, Gahmberg CG, Bianchi ME, Nawroth PP, et al. A novel pathway of HMGB1-mediated inflammatory cell recruitment that requires Mac-1-integrin. Embo J 2007;26:1129-39.
- 29) Rouhiainen A, Kuja-Panula J, Wilkman E, Pakkanen J, Stenfors J, Tuominen RK, Lepantalo M, Carpen O, Parkkinen J, Rauvala H. Regulation of monocyte migration by amphoterin (HMGB1). Blood 2004;104:1174-82.
- 30) Taniguchi N, Kawahara K, Yone K, Hashiguchi T, Yamakuchi M, Goto M, Inoue K, Yamada S,

Ijiri K, Matsunaga S, et al. High mobility group box chromosomal protein 1 plays a role in the pathogenesis of rheumatoid arthritis as a novel cytokine. Arthritis Rheum 2003; 48:971-81.

- 31) Dumitriu IE, Baruah P, Bianchi ME, Manfredi AA, Rovere-Querini P.: Requirement of HMGB1 and RAGE for the maturation of human plasmacytoid dendritic cells. Eur J Immunol 2005, 35:2184-2190.
- 32) Sharpe AH, Freeman GJ. The B7-CD28 superfamily. Nat Rev Immunol 2002;2:116-26.
- 33) Hunter RL. Overview of vaccine adjuvants: present and future. Vaccine 2002;20 Suppl 3:S7-12.
- 34)Gallucci S, Lolkema M, Matzinger P. Natural adjuvants: endogenous activators of dendritic cells. Nat Med 1999;5:1249-55.
- 35)Rock KL, Hearn A, Chen CJ, Shi Y.: Natural endogenous adjuvants. Springer Semin Immunopathol 2005;26:231-46.
- 36) Srivastava PK, Maki RG. Stress-induced proteins in immune response to cancer. Curr Top Microbiol Immunol 1991;167:109-23.
- 37) Shi Y, Evans JE, Rock KL: Molecular identification of a danger signal that alerts the immune system to dying cells. Nature 2003;425:516-21.
- 38) Rovere-Querini P, Capobianco A, Scaffidi P, Valentinis B, Catalanotti F, Giazzon M, Dumitriu IE, Muller S, Iannacone M, Traversari C, et al. HMGB1 is an endogenous immune adjuvant released by necrotic cells. EMBO Rep 2004;5:825-30.
- 39) Palumbo R, Sampaolesi M, De Marchis F, Tonlorenzi R, Colombetti S, Mondino A, Cossu G, Bianchi ME. Extracellular HMGB1, a signal of tissue damage, induces mesoangioblast migration and proliferation. J Cell Biol 2004;164:441-9.
- 40)Limana F, Germani A, Zacheo A, Kajstura J, Di Carlo A, Borsellino G, Leoni O, Palumbo R, Battistini L, Rastaldo R, et al. Exogenous highmobility group box 1 protein induces myocardial regeneration after infarction via enhanced cardiac C-kit+ cell proliferation and differentiation. Circ Res 2005;97:e73-83.
- 41)Cohen J: The immunopathogenesis of sepsis. Nature 2002, 420:885-91.

- 42) Tracey KJ, Fong Y, Hesse DG, Manogue KR, Lee AT, Kuo GC, Lowry SF, Cerami A. Anticachectin/TNF monoclonal antibodies prevent septic shock during lethal bacteraemia. Nature 1987;330:662-4.
- 43) Ohlsson K, Bjork P, Bergenfeldt M, Hageman R, Thompson RC. Interleukin-1 receptor antagonist reduces mortality from endotoxin shock. Nature 1990;348:550-2.
- 44) Reinhart K, Karzai W. Anti-tumor necrosis factor therapy in sepsis: update on clinical trials and lessons learned. Crit Care Med 2001;29:S121-5.
- 45) Fisher CJ, Jr., Dhainaut JF, Opal SM, Pribble JP, Balk RA, Slotman GJ, Iberti TJ, Rackow EC, Shapiro MJ, Greenman RL, et al. Recombinant human interleukin 1 receptor antagonist in the treatment of patients with sepsis syndrome. Results from a randomized, double-blind, placebo-controlled trial. Phase III rhIL-1ra Sepsis Syndrome Study Group. JAMA 1994;271:1836-43.
- 46) Wang H, Bloom O, Zhang M, Vishnubhakat JM, Ombrellino M, Che J, Frazier A, Yang H, Ivanova S, Borovikova L, et al. HMG-1 as a late mediator of endotoxin lethality in mice. Science 1999, 285:248-51.
- 47) Yang H, Ochani M, Li J, Qiang X, Tanovic M, Harris HE, Susarla SM, Ulloa L, Wang H, DiRaimo R, et al.: Reversing established sepsis with antagonists of endogenous high-mobility group box 1. Proc Natl Acad Sci USA 2004;101:296-301.
- 48) Hatada T, Wada H, Nobori T, Okabayashi K, Maruyama K, Abe Y, Uemoto S, Yamada S, Maruyama I. Plasma concentrations and importance of High Mobility Group Box protein in the prognosis of organ failure in patients with disseminated intravascular coagulation. Thromb Haemost 2005;94:975-9.
- 49) Wang H, Yang H, Czura CJ, Sama AE, Tracey KJ. HMGB1 as a late mediator of lethal systemic inflammation. Am J Respir Crit Care Med 2001;164:1768-73.
- 50) Wang H, Yang H, Tracey KJ. Extracellular role of HMGB1 in inflammation and sepsis. J Intern Med 2004;255:320-31.
- 51) Ulloa L, Ochani M, Yang H, Tanovic M, Halperin

D, Yang R, Czura CJ, Fink MP, Tracey KJ. Ethyl pyruvate prevents lethality in mice with established lethal sepsis and systemic inflammation. Proc Natl Acad Sci USA 2002;99: 12351-6.

- 52) Wang H, Liao H, Ochani M, Justiniani M, Lin X, Yang L, Al-Abed Y, Metz C, Miller EJ, Tracey KJ, et al. Cholinergic agonists inhibit HMGB1 release and improve survival in experimental sepsis. Nat Med 2004;10:1216-21.
- 53) Abeyama K, Stern DM, Ito Y, Kawahara K, Yoshimoto Y, Tanaka M, Uchimura T, Ida N, Yamazaki Y, Yamada S, et al. The N-terminal domain of thrombomodulin sequesters highmobility group-B1 protein, a novel antiinflammatory mechanism. J Clin Invest 2005;115:1267-74.
- 54) Abraham E, Arcaroli J, Carmody A, Wang H, Tracey KJ. HMG-1 as a mediator of acute lung inflammation. J Immunol 2000;165:2950-4.
- 55) Kokkola R, Sundberg E, Ulfgren AK, Palmblad K, Li J, Wang H, Ulloa L, Yang H, Yan XJ, Furie R, et al. High mobility group box chromosomal protein 1: a novel proinflammatory mediator in synovitis. Arthritis Rheum 2002;46:2598-603.
- 56)Kokkola R, Li J, Sundberg E, Aveberger AC, Palmblad K, Yang H, Tracey KJ, Andersson U, Harris HE. Successful treatment of collageninduced arthritis in mice and rats by targeting extracellular high mobility group box chromosomal protein 1 activity. Arthritis Rheum 2003;48:2052-8.
- 57) Jiang W, Pisetsky DS. Mechanisms of Disease: the role of high-mobility group protein 1 in the pathogenesis of inflammatory arthritis. Nat Clin Pract Rheumatol 2007;3:52-8.
- 58) Taira T, Matsuyama W, Mitsuyama H, Kawahara KI, Higashimoto I, Maruyama I, Osame M, Arimura K. Increased serum high mobility group box-1 level in Churg-Strauss syndrome. Clin Exp Immunol 2007;148:241-7.
- 59)Ito T, Kawahara K, Nakamura T, Yamada S, Abeyama K, Hashiguchi T, Maruyama I. Highmobility group box 1 protein promotes development of microvascular thrombosis in rats. J Thromb Haemost 2007;5:109-16.
- 60) Russell JA. Management of sepsis. N Engl J Med

2006;355:1699-713.

- 61)Sappington PL, Yang R, Yang H, Tracey KJ, Delude RL, Fink MP. HMGB1 B box increases the permeability of Caco-2 enterocytic monolayers and impairs intestinal barrier function in mice. Gastroenterology 2002;123:790-802.
- 62)Sappington PL, Fink ME, Yang R, Delude RL, Fink MP. Ethyl pyruvate provides durable protection against inflammation-induced intestinal epithelial barrier dysfunction. Shock 2003;20:521-8.
- 63) Saito H, Maruyama I, Shimazaki S, Yamamoto Y, Aikawa N, Ohno R, Hirayama A, Matsuda T, Asakura H, Nakashima M, et al. Efficacy and safety of recombinant human soluble thrombomodulin (ART-123) in disseminated intravascular coagulation: results of a phase III, randomized, double-blind clinical trial. J Thromb Haemost 2007;5:31-41.
- 64) Weiler H, Isermann BH. Thrombomodulin. J Thromb Haemost 2003;1:1515-24.
- 65) Esmon CT. The interactions between inflammation and coagulation. Br J Haematol 2005;131:417-30.
- 66) Riewald M, Petrovan RJ, Donner A, Mueller BM, Ruf W. Activation of endothelial cell protease activated receptor 1 by the protein C pathway. Science 2002;296:1880-2.
- 67)Bernard GR, Vincent JL, Laterre PF, LaRosa SP, Dhainaut JF, Lopez-Rodriguez A, Steingrub JS, Garber GE, Helterbrand JD, Ely EW, et al. Efficacy and safety of recombinant human activated protein C for severe sepsis. N Engl J Med 2001;344:699-709.
- 68) Conway EM, Van de Wouwer M, Pollefeyt S, Jurk K, Van Aken H, De Vriese A, Weitz JI, Weiler H, Hellings PW, Schaeffer P, et al. The lectin-like domain of thrombomodulin confers protection from neutrophil-mediated tissue damage by suppressing adhesion molecule expression via nuclear factor kappaB and mitogenactivated protein kinase pathways. J Exp Med 2002;196:565-77.
- 69) Esmon C. Do-all receptor takes on coagulation, inflammation. Nat Med 2005;11:475-7.
- 70)Levi M, Ten Cate H. Disseminated intravascular

coagulation. N Engl J Med 1999;341:586-92.

- 71)Faust SN, Levin M, Harrison OB, Goldin RD, Lockhart MS, Kondaveeti S, Laszik Z, Esmon CT, Heyderman RS. Dysfunction of endothelial protein C activation in severe meningococcal sepsis. N Engl J Med 2001;345:408-16.
- 72)Brunn GJ, Platt JL. The etiology of sepsis: turned inside out. Trends Mol Med 2006;12:10-6.
- 73)Sansonetti PJ: The innate signaling of dangers and the dangers of innate signaling. Nat Immunol 2006;7:1237-42.